

DEVELOPMENT OF HYDROXYLATED POLYCHLORINATED BIPHENYL (OH-PCBs) ANALYTICAL METHOD IN HUMAN URINE WITH UPLC/Q-TOF MS

Motoharu Suzuki¹, Toshihiro Okuno², Chisato Matsumura², Nobutake Sato³, Jun Yonekubo³, Tatsuya Ezaki³, Yoshinori Inoue⁴, Hiromasa Imaishi⁵, Takeshi Nakano^{2,6}

¹Kobe University, Graduate School of Agricultural Science, 1-1 Rokkodaicho, Nada, Kobe, Hyogo 657-8501, Japan

²Hyogo Prefectural Institute of Environmental Sciences, 3-1-27, Yukihiro-cho, Suma-ku, Kobe-city Hyogo-ken, 654-0037 Japan.

³Nihon Waters K.K., 1-3-12 Kitashinagawa, Shinagawa-ku Tokyo-to 140-0001 Japan.

⁴Nippon Filcon Co., Ltd., Products Planning & Development Dep., Research & Development Center, 2220 Ohmaru Inagi-city Tokyo, 206-8577, Japan

⁵Kobe University, Functional Analysis of Environmental Genes Research Center for Environmental Genomics, 1-1 Rokkodaicho, Nada, Kobe, Hyogo 657-8501, Japan

⁶Osaka University, Center for Advanced Science and Innovation, 2-1-1, Yamadaoka, Suita, Osaka 565-0871, Japan

Introduction

Polychlorinated biphenyls (PCBs) are known as environmental contaminants that may cause abnormal effect in various organs and some studies determined the residue levels and patterns of PCBs and metabolized PCBs (hydroxylated PCB : OH-PCBs) congeners in human blood. Hydroxyl group of OH-PCBs has high acidity, therefore OH-PCBs was made methoxy-derivatization and analyzed with GC-MS.

Derivatization GC-HRMS method has some issues that complicated preparations are needed and separation of methoxy metabolized PCBs and methoxy-derivatization OH-PCBs is difficult.

Our previous study determined elution order for 51 congeners of OH-PCB without derivatization with UPLC/QToF MS. The present study aimed at developing an analytical method for quantity to separate mixture of 6 major OH-PCBs in human blood (4'-OH-CB-107, 3-OH-CB-138, 4'-OH-CB-146, 3-OH-CB-153, 4'-OH-CB-172 and 4-OH-CB-187) and applying analytical method to biological sample of human urine.

An analytical method is developed to measure 6 major OH-PCBs in human blood without derivatization and total analytical time is within 20 minutes.

Materials and Methods

OH-PCBs Standard Solution:

Six major compounds standard solution of OH-PCBs (see Fig.1) were purchased from Wellington Laboratories Inc. (Guelph, ON, Canada) and each solution was diluted in acetone.

LC-MS/MS analysis:

Identification and quantification were performed using ultra performance liquid chromatography (UPLC: Waters Acquity UPLC system) and a high-resolution q-tof mass spectrometer (Xevo G2 QToF MS) with a resolving power of more than 20000 (Table 1)

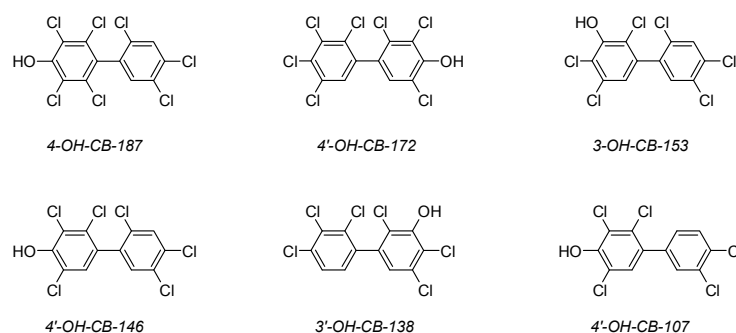


Fig.1 Major 6 components of OH-PCB in human blood

Table 1 Analytical conditions of UPLC/Q-ToF MS

<i>UPLC</i>		<i>MS</i>	
Column:	BEH C18 2.1ID X 150 mm, 1.7 μ m	ESI	negative
Flow rate	0.5 mL/min.	Capillary (kV)	1.5
Column heater	60°C	Sampling cone	40
		Source temp.	120°C
Gradient	5mM CH ₃ COONH ₄ aqTHF/CH ₃ CN (v/v: 1/4)	Desolvation temp.	600°C
Initial	75	Cone Gas Flow	20 L/hr.
17 min.	25	Desolvation Gas Flow	800 L/hr.
18 min.	1		
18.5 min.	75		

Results and Discussion

Analytical method with UPLC/Q-TOF MS is developed to separate mixture of 6 major OH-PCBs in human blood without derivatization. Figure 2 show the separation of OH-PCBs. This method is performable to separate into Six OH-PCBs. Moreover, this method enabled the separation of difficult 3-OH-153 and 4'-OH-165 in GC/HRMS with methoxy-derivatization.

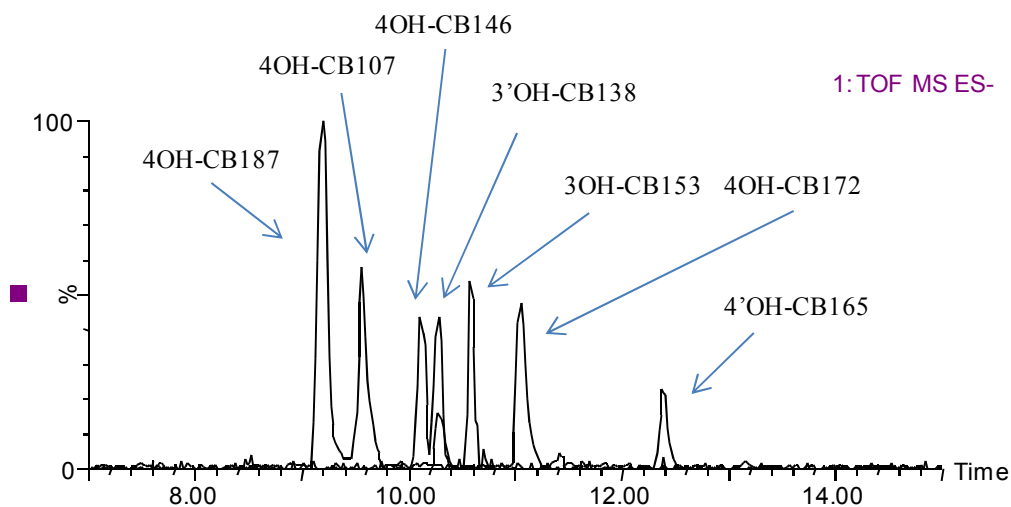


Fig2. Separation of OH-PCBs

Linearity of calibration curves at the concentration range of 0.5 to 50 ppb of 3-OH CB-138 and 4'-OH CB-153 that were detected in human urine sample is higher than 0.99 of R² (see Fig.3).

Compound name: 3-OH CB-138

Correlation coefficient: $r = 0.999553$, $r^2 = 0.999107$

Calibration curve: $51.2173 * x + -16.7275$

Response type: External Std, Area

Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis

Compound name: 4'-OH CB-153

Correlation coefficient: $r = 0.997974$, $r^2 = 0.995952$

Calibration curve: $57.0245 * x + -7.55719$

Response type: External Std, Area

Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis

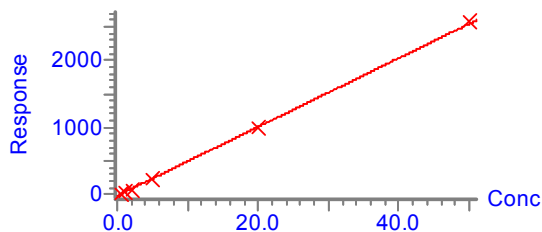


Fig3. Calibration curve of 3'OH-CB138

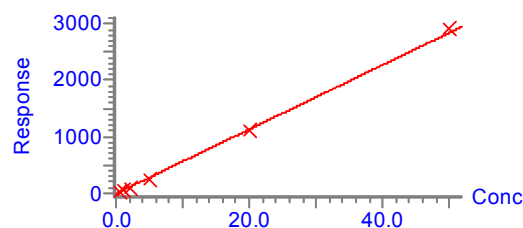


Fig.4 Calibration curve of 3OH-CB153

This method applied for human urine samples and three OH-PCBs(4OH-CB107,3'OH-CB138 and 3OH-CB153) were detected.

Table 2 Concentration of OH-PCBs in human urine.

Compounds / Samples	Urine 1	Urine 2	Urea blank
4OH-CB187	N.D.	N.D.	N.D.
4OH-CB107	4.7 pg/mL	4.5 pg/mL	N.D.
4OH-CB146	N.D.	N.D.	N.D.
3'OH-CB138	5.3 pg/mL	5.5 pg/mL	N.D.
3OH-CB153	6.0 pg/mL	9.5 pg/mL	N.D.
4OH-CB172	N.D.	N.D.	N.D.

The ratios of detected 3 OH-PCBs are show in fig.5. The order of concentration is 4OH-CB107 < 3'OH-CB138 < 3OH-CB153 in both samples.

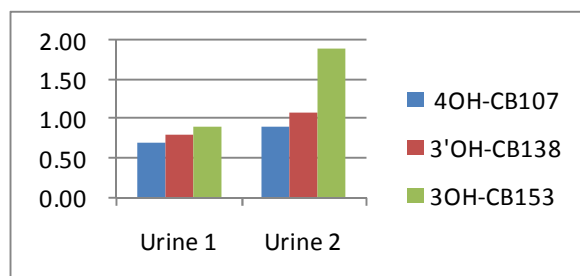


Fig.5 Ratio of detected OH-PCBs in human urine

Acknowledgements:

This research was partly supported by Grants-in-Aid for Scientific Research(B) (No. 21310027) from the Ministry of Education,Culture,Sports,Science and Technology, Japan and the Waste Management Research Grant (No. K22037) from the Ministry of the Environment, Japan.

References

- Ezaki T., Yonekubo J., Suzuki M., Matsumura C., Nakano T., 2010 Development of low level hydroxylated polychlorinated biphenyl(OH-PCBs) analytical method in human blood with UPLC/Q-Tof MS. *Dioxin* 2010
- Kuroki H., Haraguchi K., Saito H., Masuda Y., Wehler EK., Bergman A., 1993 Accumulation of hydroxylated PCB metabolites in blood. *Fukuoka Igaku Zasshi*. May;84(5), 248-56
- Britta F., Maria A., Philippe G., Pal W., Ake B., 2002. Hydroxylated PCB Metabolites and PCBs in Serum from Pregnant Faroese Women. *Environmental Health Perspectives* Vol. 110